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Distribution and Effect of Echium Plantagineum L. on Floral Species Diversity in the Dejen District, Northern Ethiopia

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ABSTRACT

Invasive plant species are considered the greatest global threat to the loss of biodiversity. Echium plantagineum L. an invasive species, is primarily found along roadsides and grazing lands in the Amhara Regional State of the Dejen district. The purpose of this study was to investigate the distribution and effect of E. plantagineum on the diversity of floral in the area of its occurrences. Reconnaissance survey was conducted to observe the distribution, spread, and effect of E. plantagineum on plant and animal species. As a result, 116 systematically established sample plots from the study were taken (58 invaded and non-invaded plots for each). To gather information on trees (shrubs) 10m x 20m sample plots were created on grazing land types. At each corner, a 1m2 subplot was taken to collect herbaceous plants. A 4m2 plot on roadside land units was created to gather information on the herbaceous flora, tree or shrub seedlings. Plants in each plot were recorded and named. Visual estimation was used to determine the percentage cover of herbaceous species. The study assessed species diversity using species evenness, Shannon Diversity Index, and Simpson Index of Dominance measures. These metrics were analyzed using PAST software. Similarity between invaded and non-invaded areas was evaluated using Jaccard's similarity index and beta diversity. The relationship between the abundance of E. plantagineum and floral species abundance was examined through linear regression and correlation analyses. A total of 85 plant species from 32 families were discovered in the study areas. In non-invaded land units, 80 plant species from 30 families were recorded. In comparison, the invaded area contained 54 plant species grouped into 24 families. Thus, the number of plants was reduced by 32.5% in the E. plantagineum-invaded area compared to the non-invaded area. The findings also showed that the non-invaded areas have a higher Shannon Diversity index value was observed in plant (3.537) and also higher plant species evenness value (0.429) compared to the E. plantagineum invaded areas. The study also showed a negative relationship between the abundance of E. plantagineum invasives and the abundance of native plant species per study plot. It was concluded that E. plantagineum, the invasive plant species in the study area, has reduced floral species diversity. Therefore, it is strongly advised to implement adequate planning and strategies to detect or stop the spread and effects of E. plantagineum. This can be achieved by creating communication channels between the regional, zonal, and district agricultural offices.

Keywords: Herbaceous species, Invaded area, Invasive plant species, Land use type, Paterson's curse

INTRODUCTION

Invasive plant species (IPSs) are non-native or alien to the ecosystem and their introduction can lead to economic or environmental harm (Byabasaija et al.). The invasion of these species is considered as the greatest global threat to biodiversity (Amare Seifu et al.). Echium plantagineum L. (Paterson's curse or Salvation Jane) is an annual or biennial species, Native to Northwest Africa, the Iberian Peninsula, and Atlantic Western Europe; it is a noxious weed in Australia. Echium plantagineum L. was first in-

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troduced in Australia as an ornamental and deliberately spread in the mid-nineteenth century (Shea et al.). It is now common, and frequently abundant throughout southwest Western Australia, central and southern South Australia, southeast Queensland, central and southern New South Wales, Victoria, Tasmania, and the Northern Territory. At the edge of its range, it is mostly limited to disturbances on roadsides. In New Zealand, E. plantagineum is abundant north of the volcanic plateau of the North Island and scattered to rare elsewhere (Ansari et al.).

It produces toxic pyrrolizidine alkaloids in the shoots (Skoneczny et al.) and bioactive shikonins in the root (Lloyd; Zhu et al.). It competes with pasture grasses and is often toxic to grazing livestock. Globally, E. plantagineum is currently infesting more than 30 million hectares of land, leading to annual losses that exceed A\$250 million (Zhu et al.). Introduced species are of global concern in terms of their inherent economic and environmental costs, with annual losses of \$1.4 trillion associated with biological invaders around the world (Bai; Hall). Echium plantagineum L. is self-incompatible in its native range but purportedly became self-compatible after its introduction to Australia (Petanidou et al.). The plant, which is present on more than 33 million hectares of Australian land (Piggin, Sheppard), is thought to harm upwards of \$30 million (AUD) annually (Shea et al.).

Invasive plant species (IPS) have significant social, ecological, and economic impacts which cause obvious changes in artificial or natural ecosystems. They pose significant challenges to agricultural lands, range lands, biodiversity, national parks, waterways, rivers, power dams, roadsides, and urban green spaces. Their presence has both economic and ecological consequences, impacting grazing areas and facilitating the spread of vector-borne diseases (Gedyon Tamiru; Wakshum Shiferaw et al.). From their threat to biodiversity and ecosystem services, invasive alien plant species have significant social, ecological, and economic impacts (Mohammed Mussa et al.).

Invasions of invasive alien plants (IAPs) serve as a hiding place for crop pests, and wild animals, and contribute to the spreading of vector-borne diseases (Mohammed Mussa et al.). Echium plantagineum L. is an alien invasive species, that may spread and grow vigorously when introduced deliberately or unintentionally by gardeners, traders, and foresters (Singh et al.) and threaten biodiversity, ecosystems, economy, and human health (Levine et al.).

Echium plantagineum L. is an annual herbaceous species, that belongs to the Boraginaceae family, it grows from 30 to 80cm in height, has rosette leaves that are up to 25cm long, and has very showy purple flowers that grow about 2 to 3cm in length arranged in an erect raceme (Weber). Africa may be principally susceptible to exotic and invasive species colonization due to its climate-sensitive distribution of native flora and fauna. In Ethiopia, IAS are causing a variety of problems for agricultural lands, roadsides, range lands, biodiversity, national parks, waterways, rivers, power dams, and urban green spaces, with serious economic and environmental significance (Gedyon Tamiru).

According to recent studies, new plant invaders that have not yet been discovered in Ethiopia are beginning to sprout, spread to uninvaded areas, and have harmful effects (Jemal Tola, Taye Tessema). While E. plantagineum was unheard of in the nation, emergent weeds like Argemone mexicana, Senna didymobotrya, and Tagetes minuta were discovered there [Figure 1].



Figure 1: Location map of the study area.

MATERIALS AND METHODS

Description of the Study Area

Geographical location: The study was conducted in the Dejen woreda (district) of the Amhara regional state in Ethiopia. The study area was located in the East Gojjam zone of the Amhara regional state, on the edge of the Abay canyon. The geographical coordinates of the study area range from approximately 10°02'28"N to 10°20'17"N latitude and 38°02'30"E to 38°18'27"E longitude. The elevation of the area varies between 1002 and 2586 m.a.s.l. It is located at a distance of 230 km from Addis Ababa, 335km from Bahir Dar by Debre Markos, and 255km connected by Motta, which is the capital city of Amhara regional state. It is bordered on the South by Abay River which separates it from the Wara Jarso woreda in the North Shoa zone of the Oromia regional state, on the West by Awabel, on the Northwest by Debay Telatgen, on the North by Enemay, and on the East by Shebel Berenta woredas of East Gojjam zone of the Amhara regional state [Figure 2].

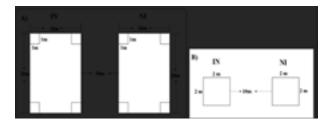


Figure 2: Layout of sample plots for plant and animal data collection (a) grazing land use type and (b) roadside land use type.

Topography and climate: According to the information provided by the Dejen District Agriculture Office, the Dejen district has a diverse landscape consisting of plains, gorges, slopes, and hilly topography. Approximately 38.8% of the district is plains (highlands), 49.4% is categorized as Weyna Dega, and 11.8% is dipping (lowland). In terms of climate, Dejen experiences a moderate climate with temperatures ranging between 17°C (63°F) and 22°C (73°F) throughout the year. The mean annual maximum temperature is 27.44°C (81.39°F), while the mean annual minimum temperature is 15.37°C (59.67°F). The mean annual average temperature is around 23°C (73°F). The district receives an average annual precipitation of about 1341 mm (52.8 in.) spread across 148 rainy days annually, with a threshold of 1 mm (0.04 in.). Dejen enjoys an average of 3897 hours of sunshine per year, and the duration of daylight varies from 11 hours 31 minutes to 12 hours 41 minutes per day.

The district has a mean annual rainfall ranging from 800 mm to 1,200 mm. Approximately 90% of the district receives an average annual rainfall of 850 to 1,000 mm. Dejen has a unimodal

rainfall pattern, with the rainy season typically extending from June to September. With an altitude range of 1,002 to 2,586 meters above sea level, the district's climate and topography make it suitable for agricultural practices. The sustainable rainfall and moderate temperatures contribute to the agricultural productivity of the area (Dejen District Agricultural Office).

Vegetation and soil type: According to the information provided by the Dejen District Agriculture Office, the district has a variety of natural resources and soil types. The natural resources include natural forests, plantation trees, shrub land, bushland, and grassland, which contribute to the ecological diversity of the area. The major soil types in the district are vertisols (clayey soils with high clay mineral content), cambisols (weathered soils), nitisols (fertile soils used for agriculture), and entisols (minimal development of soil horizons) respectively in order of proportion from high to low in terms of area coverage.

Additionally, the district is known to have various types of rocks, such as limestone, sandstone, silica sand, calcite, marble, clay soil, and granite. These geological features indicate the presence of mineral resources in the area, which could potentially support industrial sectors. Therefore, due to its diverse soil types and geological composition, Dejen district has the potential to support both agricultural activities and industrial sectors, making it suitable for a range of economic activities (Dejen District Agricultural Office).

Population: According to the 2022 national census conducted by the Central Statistical Agency of Ethiopia (CSA), Dejen woreda has a total population of 131,930. Out of this population, 63,715 are male and 68,215 are female. The woreda has a total area of 620.97 square kilometers or 62,097 hectares. With a population of 131,930, the population density of Dejen Woreda is approximately 121.5 people per square kilometer. In terms of religious affiliation, the majority of the inhabitants in Dejen woreda follow Ethiopian Orthodox Christianity, accounting for 97.01% of the population. Approximately 2.85% of the population identify as Muslims, while the remaining 0.14% follow other religions.

Farming Systems and land uses: Dejen woreda is indeed known for its agricultural activities, with a focus on both crop cultivation and livestock rearing. The agricultural production system in the district is characterized as a mixed crop-livestock farming system. The majority of the population, over 98%, derives their livelihood from agricultural practices, including crop cultivation and animal-rearing activities, and the rest 2% or less do small business.

Cattle and goat production plays a central role in the lives of many households in Dejen woreda. Livestock rearing, alongside crop cultivation, contributes significantly to the local economy and sustains the livelihoods of the community.

In addition to its agricultural significance, Dejen woreda is home to various natural resources and landmarks. These natural resources, such as forests, rivers, and fertile lands, contribute to the agricultural productivity of the area. The district's cultural and historical significance is also evident through its landmarks, which may include historical sites, traditional practices, and cultural events. Overall, the agricultural activities and natural resources in Dejen woreda play a crucial role in the lives of its residents, shaping their livelihoods and contributing to the local economy and cultural heritage of the region.

Materials

The common equipment and tools used for plant specimen collection during the study included clippers (scissors), a digger, a field press (woody frame), straps, newspapers (blotters), pencils, cameras, a meter for laying out sample plots, and GPS tools for recording latitude, longitude, and altitude. Documentation data forms were also used, recorded in a notebook.

In the field, the materials used for plant data collection included GPS, a sampling frame, a digger, scissors, plastic bags, a camera, a notebook, a pencil, and tag labels. For herbarium presses, woody frames, straps, and blotters were utilized. For collecting invertebrates, a manually made insect net was used, along with small glass bottles for storing the collected insects. Needles were used to penetrate their bodies and attach them to boards made of different materials.

Methods

Reconnaissance survey: A visit to the area was conducted to observe the distribution, spread, and effect of E. plantagineum on plant and animal species in different kebeles of the study area. The observation included assessing its presence in uncultivated areas, cultivated areas, along roadsides, grasslands (grazing land), and around construction sites. The purpose of the preliminary survey was to evaluate the distribution and level of infestation of this invasive species in various localities and land uses. The survey helped identify sites for detailed sampling of floral and faunal species and collection of these species. Based on this survey, sampling plots (quadrats) were established.

Sampling design techniques: Based on the reconnaissance survey, the study sites were purposively selected based on the level and severity of infestation by E. plantagineum. For statistically sound sampling purpose infestation along roadsides and grazing lands were selected. These two dominant land use types (grazing land and roadside) invaded by E. plantagineum were selected using a stratified random sampling method from the three selected study kebeles. The reason for selecting these two areas is that E. plantagineum is highly distributed in these areas and has a significant impact. Based on these criteria, three kebeles were selected out of 22 kebeles, and from each level of infestation site (highly, moderately, and least) one kebele was selected **[Table 1]**. For each kebele, five percent of the infested areas were sampled for both land use types (roadside and grazing land).

No	Land Use and Land Cover Type	Area in hectare	% coverage
1	Forest	3,522.51	5.7%
2	Shrubs	2,422	3.9%
3	Cultivated lands	37,023	59.6%
4	Grazing land	1,980	3.2%

Table 1: N	lajor land	l uses (Lu) and land	cover (Lc)	types in	i the Dejen d	istrict.
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5	5 Non-use		8%		
6	Others	12,143.49	19.6%		
То	tal	62,097	100%		
Source: (Dejen District Agriculture Office, 2018)					

To collect data from grazing land use types, a 10m x 20m plot was established to study tree species, shrubs. Within this plot, four 1m x 1m subplots were located at each corner to collect data on herbaceous vegetation. On roadside land use types, a 2m x 2m plot was established to gather information on herbaceous vegetation, tree or shrub seedlings, **[Figure 3]**. This plot size was chosen since the size of the roadside land units is narrow. The distribution and infestation level of E. plantagineum invasions were assessed, and the number of sample plots was determined based on the area coverage of E. plantagineum invasives. More plots were sampled from highly infested areas, while fewer sample plots were taken from least infested areas [Appendix 2].

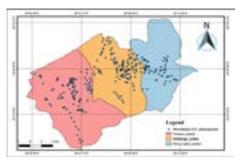


Figure 3: Distribution map of Echium plantagineum L. in three Kebeles of Dejen Woreda.

Thus, on both invaded and non-invaded roadside land use types twelve, ten, and six 2m x 2m plots (quadrats) were taken from Sebshengo, Yetnora, and Worq Amba kebeles respectively, whereas on grazing land use types (invaded and non-invaded plots) forty, thirty, and eighteen a 10m x 20m sample plots were taken from Sebshengo, Yetnora, and Worq Amba kebeles respectively [**Table 2**]. Each of the 1m x 1m subplots and 2m x 2m plots were subdivided into a 10cm x 10cm grid to measure the percentage cover for each plant species (plant cover) (Junior et al.). A Complete list of herbaceous plants was compiled for each quadrat, and the percent cover value was estimated for each species. To examine the effects of E. plantagineum invasions on the floral species diversity of invaded communities, the layout of sampling plots was standardized across all three selected kebeles. The number of sample plots was equally distributed between invaded and adjacent non-invaded areas for both land use types. As much as possible data from non-invaded plots were collected within a distance of less than 10 m from invaded plots that exhibited similar ecological and environmental gradients (Karki).

The location (latitude, longitude, and altitude) of quadrats of the study site was recorded with the help of a GPS instrument (Amare Seifu & Ermias Lulekal). A total of 116 main plots were sampled, consisting of 58 invaded plots and 58 non-invaded plots, representing both land use types. Among these, 88 quadrats (44 plots for each invaded and non-invaded) were 10m x 20m in size, representing the grazing land use type. Within each of these main plots, 352 nested plots measuring 1m2 were placed at each corner. Additionally, 28 quadrats measuring 2m x 2m were sampled for the roadside land use type [Table 3] [Appendix 2].

Data collection: Data collection was conducted between January and February 2025 to assess the effect of E. plantagineum on floral species diversity in different infested land use types. Each native plant species, invasive plant species were identified and recorded. The percentage cover of the invasive and herbaceous species was visually estimated by subdividing plots into a 10cm x 10cm grid (Bibby).

Plant species discovered in the quadrats were identified in the field. Sample specimens of the recorded plant species were collected, pressed, properly dried, and taken to the Ethiopian Biodiversity Institute Herbarium for identification following (Amare Seifu & Ermias Lulekal). The collected specimens were identified by comparing them with authenticated specimens, consulting experts, and referring to the published volumes of Flora of Ethiopia and Eritrea (Hedberg, 1996; Mesfin Tadesse).

Data analysis:

Descriptive (frequency and percentage) and inferential statisti-

Study kebeles	Level of infestation	No of sample plots		
		Roadside	Grazing land	Total
Sebshengo	Highly infested	12 (6 IN & 6 NI)	40 (20 IN & 20 NI)	52
Yetnora	Moderately infested	10 (5 IN & 5 NI)	30 (15 IN & 15 NI)	40
Worq Amba	Least infested	6 (3 IN & 3 NI)	18 (9 IN & 9 NI)	24
Т	otal	28 (14 IN & 14 NI)	88 (44 IN & 44 NI)	116

Table 2: Numbers of quadrats for selected study sites (kebeles).

Table 3: Effect of Echium plantagineum L. on herbaceous and woody plant diversity, richness, and evenness.

No	Parameters	Non-invaded	Invaded	% reduction over non-invaded
1	Richness (total species)	80	54	(-) 32.5%
2	No. of Individuals	832	552	(-) 33.65%
3	Dominance_D	0.047	0.578	(+) 91.87%

4	Simpson_1-D	0.953	0.422	(-) 55.72%	
5	Shannon_H'	3.537	1.389	(-) 60.73%	
6	Evenness_e^H/S	0.429	0.073	(-) 83%	
7	Jaccard's similarity index	0.			
8	Beta diversity	0.42			
(-) sign show less value and (+) sign show high value in invaded site					

cal tools were used to present, interpret, and analyze the data. To determine the distribution of E. plantagineum invasive species, the study utilized QGIS 3.18.3 software. In this study, GPS coordinates or spatial polygons data indicating the invaded sites on the occurrence or presence of E. plantagineum at different locations within the study area was gathered. Using QGIS, this spatial data has imported into the software and georeferenced it to align with the study area's underlying base map or reference system. This software was employed to generate a distribution map, providing visual representation and analysis of the invasive species' spread and presence in the study area. The biophysical data, including the total number of plant species, and number of individuals were organized using Microsoft Office Excel. The data collected from the plot survey were analyzed using different statistical tools with the help of IBM SPSS (Statistical Package for Social Sciences) Statistics version 26 software.

In the study area, the diversity of species in E. plantagineum-infested sample sites was evaluated using several metrics, including the Shannon Diversity index (H'), species evenness (E), Dominance Index (D), and Simpson Index of Dominance (1-D). These metrics were analyzed using PAST software, which allowed for statistical analysis and interpretation of the diversity indices in relation to the presence and effect of E. plantagineum. The purpose was to evaluate the impact of E. plantagineum on the growth and distribution of other plant species.

The species evenness value ranges between 0 and 1. A value of 0 indicates that a few species are highly abundant, while a value of 1 signifies that all species are equally abundant in the sample. The index of dominance values range between 0 and 1. A value closer to 0 indicates a more diverse plot, while a value closer to 1 suggests that the plot is dominated by a few species. The pattern observed for dominance index values is opposite to that of Shannon index values. As the Shannon Diversity value increases, the Index of dominance value decreases.

To compare infested and non-infested areas Jaccard's similarity index and beta diversity were calculated (Musese et al.). The frequencies of each species in non-invaded and invaded sites were determined. Jaccard's similarity index and Sorensen's similarity index were computed to assess the similarity between non-invaded and invaded areas for the selected land use types in each sampling site (Karki).

Jaccard's Similarity Index (JSI) was calculated as JSI=c/(a+b-c), where c represents the number of species common to both infested and non-infested areas, b denotes the species present in infested areas but absent in non-infested areas, and a represents the species present in non-infested areas but absent in infested areas. Alternatively, Jaccard's similarity index can be calculated as JSI=c/(a+b-c), where c is the number of species shared by in-

vaded and non-invaded communities, a is the number of species in the invaded community, and b is the number of species in the non-invaded community. The coefficient ranges from 0 to 1, where a value of 1 indicates complete similarity and a value of 0 signifies complete dissimilarity between the compared areas.

According to Amare Seifu et al., the beta diversity index was calculated to assess the impact of invasion on species composition and the relative covers of resident species in invaded and non-invaded study sites. The calculation of the Beta diversity index was based on species covers (Hejda et al.). It is calculated using the formula β =1-JSI, where JSI is Jaccard's similarity index (Baselga).

A linear regression and correlation analysis was conducted to evaluate the relationship between the abundance of E. plantagineum and the abundance of floral and species within study quadrats. The regression equation was computed as, y=a+bx, where y represents the dependent variable (parameters), a is the regression constant, b is the estimated regression coefficient, and x represents the independent variable (parameter). In this case, the independent variable (x) is the abundance of E. plantagineum, while the dependent variable (y) is the abundance of specific dominant non-invasive species.

RESULTS AND DISCUSSION

Distribution of Echium plantagineum L. in the Study Area

Echium plantagineum L. has been observed to grow on roadsides, grazing land, and occasionally on poorly tilled farmlands. Among these habitats, roadside and grazing land use types are frequently infested by this invasive weed [Appendix 1]. The high frequency of infestation along roadsides can be attributed to continuous disturbance and the transportation of sand and soil during road construction and maintenance. The distribution of E. plantagineum is expanding rapidly in the research area due to its prolific seed production, posing a significant threat to roadsides and grazing lands.

A similar study was conducted on the abundance and geographical distribution of invasive weed species in the Western Amhara Region, Ethiopia, by Melkamu Birhanie et al. (2020), who reported that the distribution of E. plantagineum is escalating at an alarming rate and invading roadsides and grazing lands along the Dejen to Debre Markos Road. Among the three selected study kebeles, Sebshengo kebele exhibited the widest distribution of E. plantagineum compared to the other two kebeles [Figure 4]. The invasive species has covered substantial portion of the areas within the kebeles. In contrast, Worq Amba kebele showed the least colonization by E. plantagineum invasives [Figure 4].

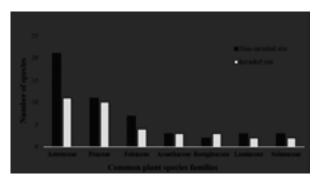


Figure 4: Comparison of plant families between invaded and non-invaded plots.

Effect of Echium plantagineum L. on Plant Species Diversity

In the non-invaded area, there were a record of 80 plant species belonging to 30 families. Whereas, in the E. plantagineum invaded sites, there were a record of 54 plant species belonging to 24 families [Appendix 3]. Among 85 plant species, 49 species were found common in non-invaded and invaded areas. Thirty-one plant species (Acacia mearnsii De Wild., Achyranthes aspera L., Acmella caulirhiza Del., Argemone mexicana L., Artemisia ludoviciana Nutt., Calpurina aurea (Ait.) Benth., Cirsium vulgare (Savi) Ten., Clematis hirsuta Perr. & Guill., Crepis dioscoridis L., Cucumis ficifolius A.Rich, Desmodium intortum (Mill.) Urb., Dodonaea viscosa (L.) Jacq., Erigeron bonariensis L., Ficus palmata Forssk., Laggera crispata (Vahl) Hepper & J.R.I.Wood., Lippia adoensis Hochst. ex Walp., Leucas martinicensis (Jacq.) R.Br., Nuxia congesta Fresen., Osyris lanceolata Hochst. & Steud., Premna schimperi Engl., Rhus vulgaris Meikle, Rhynchosia ferruginea A. Rich., Rosa abyssinica Lindley, Sonchus asper (L.) Hill, Solanum stramoniifolium Jacq., Sphaeranthus suaveolens (Forssk) DC., Stephania abyssinica (Dill. & Rich.) Walp., Tagetes minuta L., Verbascum benthamianum L., Verbena officinalis L., and Vernonia amygdalina Del.) were found only in non-invaded areas. While five species (Cupressus lusitanica Mill., Echium plantagineum L., Euphorbia candelabrum Trema., Sesbania sesban (L.) Merr., and Maytenus arbutifolia. (A. Rich) Wilezek.) were growing only in the invaded areas.

Thus, the number of species and the number of families were reduced by 32.5% and 20% respectively in the E. plantagineum-invaded area as compared to the non-invaded area. In addition, the number of individuals also reduced by 33.65% in the invaded area compared to non-invaded. The decline in plant species and families indicates that the E. plantagineum invasion could have resulted in a reduction in plant diversity and a change in the composition of plant communities in the invaded sites. The results of the study indicate that the invasion of E. plantagineum had an impact on the diversity and composition of flora in the study area.

The Shannon Diversity index value for non-invaded plots was 3.537, while for invaded plots it was 1.389. Similarly, the species evenness index value for non-invaded plots was 0.429, while for invaded plots it was 0.073. Thus, Shannon's index of diversity and evenness index value were reduced by 60.73% and 83% respectively in the E. plantagineum invaded areas compared to non-invaded plots [Table 4]. These findings indicate that the invasion of E. plantagineum has resulted in a decrease in herbaceous and woody plant species richness, diversity, and evenness. These results showed that the non-invaded areas have a higher Shannon Diversity index (3.537) and higher species evenness value (0.429) compared to the E. plantagineum invaded areas. The lower values of Shannon's index of diversity (1.389) and lower evenness index value (0.073) suggest that the invaded communities have a lower variety of species. The distribution of individuals among species is uneven, with one or a few species dominating the invaded areas. Evenness has values between 0 and 1, where 1 represents a situation in which all species are equally abundant.

Correlations		C. dactylon	A. distachyos	C. longus
E. plantagineum	Pearson Correlation	291**	319**	251**
	Sig. (2-tailed)	.002	.000	.007
	N	116	116	116
**. Correlation is significant at the 0.01 level (2-tailed), and N – No of plots.				

Table 4: Pearson correlations between the abundance of Echium plantagineum L. and plant species abundance

This finding is similar to the findings of studies by Dogra et al., Amare Seifu et al., Byabasaija et al. and Amare Seifu, Ermias Lulekal, which found that the lower values of Shannon's index of diversity (H'), and evenness index value in the invaded areas. In contrast, the higher values of Shannon's index of diversity and evenness index in non-invaded communities indicate greater species heterogeneity and a more balanced distribution of individuals among different species. Accordingly, a lower value of the Shannon Diversity index suggests an area was dominated by a few species, i.e., invaded sites were relatively homogeneous species in the community. Therefore, in the non-invaded study sites, the communities were more diverse in the type of species and heterogeneous in the community.

According to Barnes, invaded communities have low diversity because invasive plants alter the invaded ecosystem and species

composition to such an extent that they threaten native flora. Thus, the higher value of the Shannon Diversity index in the non-invaded plots suggests that E. plantagineum has an impact on the indigenous species composition through a decrease in species diversity. This harmonizes with many studies conducted in different parts and vegetation types of the world (Hailu Shiferaw et al.; Melkamu Kifetew, Zerihun Woldu), which have reported that in natural areas the shrub has a series of deleterious effect on some plant species and is known to displace natural bush communities as well as prevent natural regeneration of some tree species.

The Jaccard's Similarity Index of the communities in the invaded and non-invaded areas was 58% (similarity index), which showed loss of species was due to the invasion of E. plantagineum and that of the beta diversity index (dissimilarity index) was 42%. This indicated that E. plantagineum invaded site is dominated by a limited number of species. The uneven distribution of individuals among species in the invaded areas suggests that only a small number of species are well-adapted to the environment altered by E. plantagineum invasion. If those few species that can survive are extremely affected, then the site that contains the species may be disturbed as compared to non-invaded areas.

This finding is consistent with the Amare Seifu et al. study, which reported that a low diversity index value suggests an area is dominated by a limited number of plant species. The dissimilarity values showed significant differences between E. plantagineum invaded and non-invaded plots. Similarly, this finding is in agreement with Kohli et al. and Mesfin Boja et al., who reported fact that plant species diversity in the non-invaded area was greater than in the E. plantagineum-invaded areas. Furthermore, the index of dominance (D) value was found to have increased by 91.87% in the invaded areas compared to the non-invaded areas **[Table 4]**. The increase in dominance indicates that a smaller number of plant species have become more dominant in the in-vaded areas, potentially leading to a shift in the composition and structure of the plant community.

This finding is in line with the studies by Dogra et al., Wambua, Gebrehiwot, Berhanu, Amare Seifu, Ermias Lulekal, and Melkamu Kifetew, Zerihun Woldu who found that a higher index of dominance value in invaded areas and a lower index of dominance value for floral communities in non-invaded areas would indicate that the invaded communities were homogenous whereas non-invaded floral communities would be indicated that the heterogeneous species nature. These results collectively highlight the negative impact of E. plantagineum invasion on plant diversity and community dynamics in both herbaceous and woody vegetation.

Effect of Echium plantagineum L. on plant species composition: The results of this study showed that the invasion of E. plantagineum affected the composition of woody and herbaceous plant species in the study area. At the family level, in the non-invaded areas, of the 30 families, the family Asteraceae represented the highest number of species (21 species), accounting for 26.3%, followed by Poaceae (13.8%), and Fabaceae (8.8%). It is worth noting that the above-mentioned families alone represent 48.9% of plant species in the total flora in non-invaded study sites [Appendix 3]. Whereas, in the invaded areas, of the 24 plant families, the Asteraceae family was the highest-ranking in the number of species (11 species) accounting for 20.4%, followed by Poaceae (18.5%), and Fabaceae (7.4%). The above-mentioned three families shared 46.3% of plant species recorded in the E. plantagineum-invaded study areas [Appendix 3].

The current study indicated that the three families Asteraceae, Poaceae, and Fabaceae were the dominant families for the whole identified plant species for the study sites **[Figure 5]**. This finding is in agreement with Tsegay Gebregergs, who studied herbaceous species composition which indicated that the three families Poaceae, Asteraceae, and Fabaceae took the lead of the herbaceous species sampled. Further studies by Lema Etefa, Genet Atsbeha et al., Amare Seifu et al., and Bhatta et al. on the floristic composition of herbaceous flowering plant species found the three families Poaceae, Asteraceae, and Fabaceae have the highest number of species.

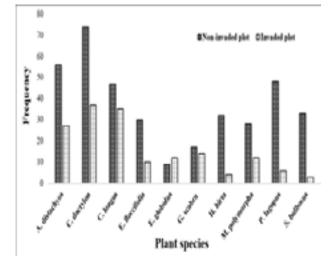


Figure 5: Comparison of some frequently recorded plant species between invaded and non-invaded plots.

Out of the 80 plant species recorded from non-invaded plots, Cynodon dactylon had the highest frequency, followed by Andropogon distachyos, Plantago lagopus, and Cyperus longus **[Figure 6]**. On the other hand, out of the 54 plant species recorded from invaded plots, Echium plantagineum had the highest frequency, followed by Cynodon dactylon, Cyperus longus, and Andropogon distachyos **[Figure 6]**.

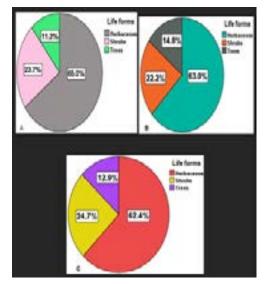


Figure 6: Different life forms of plant species recorded (a) non-in-vaded plot, (b) invaded plot, and (c) entire plot.

Regarding to growth habits, among 80 plant species identified from non-invaded plots, the majority were herbaceous species (52 species), followed by shrub species (19), and nine tree species. Whereas, among the 54 plant species identified from invaded plots, 34 species were herbaceous, followed by shrub species (12), and eight tree species. Overall, a total of 85 documented plant species, including 53 herbaceous species, 21 shrub species, and 11 tree species, were recorded from the entire study sites of both land use types **[Figure 7]**.

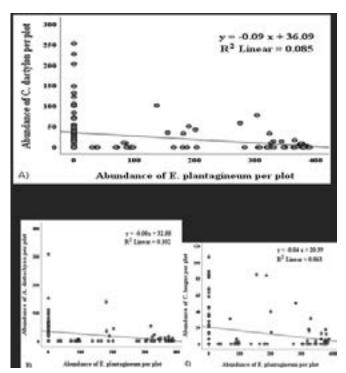


Figure 7: Relationship between the abundance of E. plantagineum and abundance of plant species (a) abundance of E. plantagineum Vs abundance of C. dactylon (b) abundance of E. plantagineum Vs abundance of A. distachyos and (c) abundance of E. plantagineum Vs abundance of C. longus per plot.

A pie chart **Figure 7** depicted that the majority of plant species recorded from both invaded and non-invaded plots were herbaceous species, surpassing the number of shrub or tree species.

Relationship between the abundance of Echium plantagineum L. and the abundance of non-invasive plant species: In all of the research plots, Cynodon dactylon, Andropogon distachyos, and Cyperus longus were the three most abundant native plant species. A scatter plot revealed that an increase in the abundance of E. plantagineum led to a decrease in plant species' abundance of native flora. The results indicated that there was a negative linear relationship between the abundance of E. plantagineum and the abundance of C. dactylon species in each study plot. Thus, the regression equation can be presented as $y=-0.09 \times x+36.06$, where y is an abundance of C. dactylon and x is the abundance of E. plantagineum per plot. In this case, R2=0.085, or 8.5% indicating that there is a weak negative relationship between the abundance of E. plantagineum and the abundance of C. dactylon plant species. The regression equation and Pearson correlation (r=-0.291, p=0.002) at p=0.01 indicated the existence of a negative linear relationship between the abundance of E. plantagineum and the abundance of C. dactylon species in each plot. The regression analysis also showed a negative relationship between the abundance of E. plantagineum and C. longus species abundance per plot. Hence, the regression equation can be presented as y=-0.04 x+20.39, where y is C. longus abundance and x is E. plantagineum abundance per plot. The regression equation indicated that there was a weak negative relationship between the abundance of E. plantagineum and C. longus species abundance. As the abundance of E. plantagineum increased species abundance of C. longus decreased. In this case, R2=0.063 or 6.3% of the dependent variable (C. longus species abundance) can be explained by

the independent variable (abundance of E. plantagineum). The regression equation and Pearson correlation (r=-0.251, p=0.007) at p=0.01 indicated the existence of a negative linear relationship between the abundance of E. plantagineum and the abundance of C. longus species per plot.

All scatter plots showed that there was a negative linear relationship between the abundance of E. plantagineum and the abundance of native plants. The regression analyses revealed that as the abundance of E. plantagineum increased, the abundance of C. dactylon, A. distachyos, and C. longus decreased. The R2 values indicated that only a small percentage of the variation in native plant abundance could be explained by the abundance of E. plantagineum. Overall, these findings suggest that E. plantagineum has a negative influence on the abundance of native plant species in the studied plots.

The results showed that there was also a negative linear relationship between the abundance of E. plantagineum and the abundance of A. distachyos plant species in the study plot. Thus, the regression equation can be written as y=-0.08 x+32.88, where y represents the abundance of A. distachyos and x is the abundance of E. plantagineum per plot. In this case, R2=0.102, or 10.2% indicated that there is a weak negative relationship between the abundance of E. plantagineum and the abundance of A. distachyos plant species. The regression equation and Pearson correlation (r=-0.319, p=0.00) at p=0.01 indicated the existence of a negative linear relationship between the abundance of E. plantagineum and the abundance of A. distachyos species per plot.

Multiple studies consistently demonstrate that invasive plants have negative effects on native plant communities. These effects include reductions in abundance, diversity, richness, and evenness of native plant species. The findings from Rejmanek, Richardson, D'Antonio, Meyerson, Daehler, Vilà et al., Catford et al., and Pyšek et al. collectively support the notion that invasive plants outcompete native species for resources, leading to declines in native plant abundance and diversity. Specifically, observed the negative relationship between the abundance of the invasive plant species E. plantagineum and the abundance of native plant species C. longus, C. dactylon, and A. distachyos suggests that the presence of E. plantagineum may indeed negatively affect the abundance of these native plants.

CONCLUSION AND RECOMMENDA-TIONS

Conclusion

Echium plantagineum L. was primarily observed in Yetnora Kebele, which serves as the location for the Kajima Road construction contractor camp. It is concerning that its distribution is rapidly increasing, invading roadsides and grazing lands within the study area. The high frequency of its presence along roadsides can be attributed to road construction and maintenance activities. The result of the study indicates that E. plantagineum is the dominant invasive alien plant species (IAPS) in the selected study Kebeles within the study area. Its presence has a significant impact on the diversity and composition of both plant and animal species in the invaded communities. The vegetation in invaded areas showed lower species richness and evenness of herbaceous and woody plant species compared to non-infested vegetation in both land use types. Further, the infestation of invasive also had negative effects on animal species diversity and composition. This finding reported that higher diversity indices were relatively recorded in the non-invaded areas in all study sites when compared to invaded areas. The study findings indicated a negative relationship between the abundance of E. plantagineum and the abundance of faunal and floral species diversity. This suggests that as the abundance of E. plantagineum increased, the diversity of faunal and floral species decreased. Overall, these findings highlight the negative effects of E. plantagineum on the diversity and composition of both plant and animal species in the invaded areas.

Recommendations

Based on the results obtained from the study, several recommendations were put forward:

- Proper management strategies and control measures are necessary to prevent further spread and mitigate its negative ecological and economic consequences.
- Create public awareness about the negative impacts of E. plantagineum on plant and animal productivity, and biodiversity, especially among farmers. This will aid in timely and effective management, preventing further spread into new areas.
- Identify priority kebeles in the district to control the spread of E. plantagineum. Targeted measures can be implemented in these areas to prevent its expansion.
- Protect and restore sensitive land use types through an integrated approach, considering ecological needs. Various strategies should be employed to manage E. plantagineum in these areas.
- Conduct further studies to assess the impacts on crop yield, human and animal health, and biodiversity. These studies will inform management practices in the study area.
- Establish communication links between agricultural offices and farmers at different levels to detect and eradicate the spread and negative impacts of E. plantagineum. This coordination will enable effective response and management.

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None.

CONFLICT OF INTEREST

None.

REFERENCES

- 1. Aguero JI, Pérez-Méndez N, Torretta JP, et al. (2020) Impact of Invasive Bees on Plant-Pollinator Interactions and Reproductive Success of Plant Species in Mixed Nothofagus antarctica Forests. Neotropical Entomology. 49(4):557-567.
- 2. Amare S, Ermias L. (2021) Impact of invasive alien plant species, Lantana camara, on the species composition, richness and evenness of invaded plant communities in Sidama, Gamogofa and Gedio Zones, Ethiopia. 1:1-34.
- 3. Amare S, Nigussie S, Manaye M, et al. (2017) Impact of Invasive Alien Plant, Xanthium Strumarium, On Species Diversity and Composition of Invaded Plant Communities in Borena Zone, Ethiopia. Biodiv Internat J. 1(1):15-22.
- 4. Ansari AA, Gill SS, Abbas ZK, et al. (2016) Plant Biodiversity:

© Under License of Creative Commons Attribution 4.0 License

Monitoring, Assessment and Conservation. CABI. 628p.

- Bai BB. (2014) Biological invasions: Economic and environmental costs of alien plant, animal, and microbe species. 37(1):277.
- 6. Barnes BV. (1998) Forest Ecology. 4th Edition. Wiley. University of Michigan. 774p.
- Baselga A. (2010) Partitioning the turnover and nestedness components of beta diversity. Global Eco Biogeo. 19(1):134-143.
- Bell LW, Lemerle D, Kirkegaard JA, et al. (2016) Benefits of break crops and weeds to wheat in southern Australia: variation with rainfall, soil type and location. Crop Pasture Sci. 67(5):497-514.
- 9. Berti M, Johnson BL, Dash S, et al. (2007) Echium: A Source of Stearidonic Acid Adapted to the Northern Great Plains in the US. 6:120-125.
- 10. Bhatta S, Joshi LR, Shrestha BB. (2020) Distribution and impact of invasive alien plant species in Bardia National Park, western Nepal. Environ Conser. 47(3):197-205.
- 11. Bibby CJ, Collar NJ, Crosby MJ, et al. (1992) Putting biodiversity on the map: priority areas for global conservation. Internat Council Bird Preser. 333(95):993.
- 12. Boy G, Witt A. (2013) Invasive alien plants and their management in Africa. Synthesis Report of the UNEP/GEF Removing Barriers to Invasive Plant Management in Africa (RBIPMA) Project. Sarah Hilliar, CABI. 181p.
- 13. Broster J, Koetz E, Wu H. (2012) Weed species present in cereal crops in southern New South Wales. 244 p.
- 14. Byabasaija S, Natasha R, Eunice C, et al. (2020) Abundance, distribution and ecological impacts of invasive plant species in Maputo Special Reserve, Mozambique. Internat J Biodiv Conserv. 12(4):305-315.
- 15. Catford JA, Daehler CC, Murphy HT, et al. (2012) The intermediate disturbance hypothesis and plant invasions: Implications for species richness and management. Persp Plant Eco, Evol Systema. 14(3):231-241.
- 16. Clusella-Trullas S, Garcia RA. (2017) Impacts of invasive plants on animal diversity in South Africa: A synthesis. Bo-thalia. 47(2):12.
- 17. Colegate SM, Edgar JA, Knill AM, et al. (2005) Solid-phase extraction and HPLC-MS profiling of pyrrolizidine alkaloids and their N-oxides: A case study of Echium plantagineum. Phytochem Anal. 16(2):108-119.
- Corbet SA, Delfosse ES. (1984) Honeybees and the nectar of Echium plantagineum L. in southeastern Australia. Austral Eco. 9(2):125-139.
- Daehler CC. (2003) Performance Comparisons of Co-Occurring Native and Alien Invasive Plants: Implications for Conservation and Restoration. Ann Rev Eco, Evol, Systema. 3:183-211.
- 20. D'Antonio C, Meyerson LA. (2002) Exotic Plant Species as Problems and Solutions in Ecological Restoration: A Synthesis. Restoration Eco. 10(4):703-713.
- 21. Das D, Banerjee S, John R. (2019) Predicting the distribution and abundance of invasive plant species in a sub-tropical woodland-grassland ecosystem in northeastern India. Plant Eco. 220(10):935-950.
- 22. Davis AR. (1992) Evaluating honey bees as pollinators of virgin flowers of Echium plantagineum L. (Boraginaceae) by

pollen tube fluorescence. J Apicul Res. 31(2):83-95.

- 23. Dellow J, Seaman J. (1985) Association with pyrrolizidine alkaloid poisoning in horses in New South Wales. 1(2):79-83.
- Dellow J, Wilson G, King W, et al. (2002) Occurrence of weeds in the perennial pasture zone of New South Wales. Plant Protec Quart. 17(1):12-16.
- 25. Department of Jobs PR. (2020) Paterson's curse Agriculture (Victoria). Agriculture Victoria. https://agriculture.vic.gov. au/biosecurity/weeds/weeds-information/patersons-curse
- Dogra KS, Kohli RK, Sood SK, et al. (2009) Impact of Ageratum conyzoides L. on the diversity and composition of vegetation in the Shivalik hills of Himachal Pradesh (Northwestern Himalaya), India. Internat J Biodiv Conserv. 1(5):135-145.
- 27. Eberle CA, Forcella F, Gesch R, et al. (2014) Flowering Dynamics and Pollinator Visitation of Oilseed Echium (Echium plantagineum). PLoS ONE. 9(11):e113556.
- El-Shazly A, Wink M. (2014) Diversity of pyrrolizidine alkaloids in the Boraginaceae structures, distribution, and biological properties. Diversity. 6(2):188-282.
- 29. Florentine S, Weller S, King A, et al. (2018) Seed germination response of a noxious agricultural weed Echium plantagineum to temperature, light, pH, drought stress, salinity, heat and smoke. Crop Pasture Sci. 69(3):326-333.
- Gebrehiwot N, Berhanu L. (2015) Impact of parthenium on species diversity in Gamo Gofa, Ethiopia. Schol J Agri Sci. 5(7):226-231.
- Tamiru G. (2017) Invasive Alien Weed Species Distribution, Impacts on Agriculture, Challenge and Reaction in Ethiopia: A Review. J Biol. 7(7):136-146.
- Atsbeha G, Demissew S, Woldu Z, et al. (2015) Floristic composition of herbaceous flowering plant species in Lalay and Tahtay Michew Districts, Central Zone of Tigray, Ethiopia. Afr J Ecol Ecosyst. 2(6):159-169.
- 33. Goulson D, Hanley ME, Darvill B, et al. (2002) Causes of rarity in bumblebees. Biol Conserv. 103(2):283-291.
- Government of Canada, CFIA. (2014) Appendix 7A RMD-13-04: Consolidated Pest Risk Management Document for pest plants regulated by Canada.
- 35. Groves RH, Boden R, Lonsdale M. (2005) Jumping the Garden Fence: Invasive Garden Plants in Australia and their environmental and agricultural impacts. CSIRO report for WWF-Australia, Sydney. 173.
- Groves RH, Panetta FD. (2014) The biology of Australian weeds - A short history of the series. Plant Prot Q. 29(4):127-130.
- 37. Shiferaw H, Teketay D, Nemomissa S, et al. (2003) Some biological characteristics that foster the invasion of Prosopis juliflora at Middle Awash Rift Valley Area, NE Ethiopia. J Arid Environ. 58(2):134-153.
- Hall CM. (2019) Biological invasion, biosecurity, tourism, and globalisation. In: Handbook of Globalisation and Tourism. 114-125.
- Hansen MJ, Clevenger AP. (2005) Influence of disturbance and habitat on non-native plant species along transport corridors. Biol Conserv. 125(2):249-259.
- Hedberg I. (1996) Flora of Ethiopia and Eritrea. In: van der Maesen LJG et al. (eds) The Biodiversity of African Plants. Springer. 802-804.
- 41. Hejda M, Pyšek P, Jarošík V. (2009) Impact of invasive plants

on species richness and composition of invaded communities. J Ecol. 97(3):393-403.

- 42. Hu Y, Jiang Z, Leung KSY, et al. (2006) Determination of naphthoquinones in Boraginaceous herbs by HPLC. Anal Chim Acta. 577(1):26-31.
- 43. Hulting AG, Krenz JE, Parker R. (2007) Paterson's curse (Echium plantagineum) in the Pacific Northwest. 4.
- 44. Huxtable RJ. (2019) Toxicology of alkaloids in foods and herbs. In: Food Poisoning. Routledge. pp. 237-262.
- 45. Tola J, Tessema T. (2015) Abundance and distribution of invasive alien plant species in Illu Ababora Zone, Oromia, Ethiopia. J Agric Sci Food Technol. 1(7):94-100.
- 46. Junior NDO, Diniz ES, Santos AR Jr. (2022) Phylogenetic structure of Pampean Grasslands under alien species. Preprints.
- 47. Karki D. (2009) Ecological and socio-economic impacts of Parthenium hysterophorus invasion in urban Nepal. MSc Thesis Tribhuvan Univ Kathmandu. 92.
- Kicel A, Piotrowska-Seget Z, Różańska A. (2018) Echium plantagineum: Phytochemical properties and medicinal perspectives. Int J Mol Sci. 19(3):708.
- 49. Kilawe CJ, Mchelu HA, Emily CJ. (2022) Impact of Cedrela odorata on Lygodactylus williamsi in Kimboza Forest, Tanzania. Glob Ecol Conserv. 38:e02225.
- Kohli RK, Dogra KS, Batish DR, et al. (2004) Impact of invasive plants on NW Indian Himalayas vegetation. Weed Technol. 18(1):1296-1300.
- Konarzewski TK. (2012) Clinal variation in life-history traits of the invasive plant species Echium plantagineum L. PhD Thesis Univ of Technology Sydney. 179.
- 52. Kovács-Hostyánszki A, Szigeti V, Miholcsa Z, et al. (2022) Threats and benefits of invasive alien plant species on pollinators. Basic Appl Ecol. 64:89-102.
- 53. Kumagai Y, Shinkai Y, Miura T, et al. (2012) Chemical biology of naphthoquinones and environmental implications. Annu Rev Pharmacol Toxicol. 52(1):221-247.
- 54. Latif S, Gurusinghe S, Weston LA. (2022) Potential role of allelopathy in invasive weeds. In: Persistence Strategies of Weeds. 271-301.
- 55. Etefa L. (2011) Floristic composition and diversity of herbaceous flowering plants in Menagesha Suba State Forest, Oromia, Ethiopia. MSc Thesis. Addis Ababa Univ., Ethiopia. 78.
- Levine JM, Vilà M, Antonio CMD, et al. (2003) Mechanisms underlying the impacts of exotic plant invasions. Proc R Soc Lond B Biol Sci. 270(1517):775-781.
- 57. Litt AR, Cord EE, Fulbright TE, et al. (2014) Effects of invasive plants on arthropods. Conserv Biol. 28(6):1532-1549.
- 58. Lloyd S. (2011) Weed management in a changing world: Acknowledgments and reviews. 1:1-609.
- 59. Loh ZH, Ouwerkerk D, Klieve AV, et al. (2020) Toxin degradation by rumen microorganisms: A review. Toxins. 12(10):664.
- 60. Martinello M, Borin A, Stella R, et al. (2017) QuEChERS method for pyrrolizidine alkaloids in honey. Food Chem. 234:295-302.
- 61. Matthews PA, Baldo BA, Howden MEH. (1988) Cytochrome c allergens in pollens of Plantago lanceolata and Echium plantagineum. Mol Immunol. 25(1):63-68.
- 62. McCulloch GA, Mauda EV, Chari LD, et al. (2020) Genetic diversity and variation in African boxthorn (Lycium ferocissimum). Biol Control. 143:104206.

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- 63. Mehrabani M, Ghannadi A, Sajjadi E, et al. (2006) Toxic pyrrolizidine alkaloids of Echium amoenum. DARU J Pharm Sci. 14(3):122-127.
- 64. Birhanie M, Zegeye W, Asaye G, et al. (2020) Abundance and distribution of invasive weeds in Western Amhara, Ethiopia. 5(1):18-25.
- 65. Kifetew M, Woldu Z. (2021) Impact of Lantana camara on native species and socio-economy near Adama and Bishoftu, Ethiopia. J Environ Earth Sci. 11(3):29-42.
- 66. Boja M, Girma Z, Dalle G. (2022) Impacts of Parthenium hysterophorus on plant diversity in Ginir District, Ethiopia. Diversity. 14(8):675.
- 67. Tadesse M. (2004) Flora of Ethiopia and Eritrea. Asteraceae. 4(2):408.
- 68. Moerkerk M. (2006) Risk of weed movement via vehicles and equipment: A Victorian study. 15th Aust Weeds Conf. 4.
- 69. Mussa M, Teka H, Aliye A. (2018) Socio-economic and environmental impacts of invasive plants in Bale Zone, SE Ethiopia. Afr J Agric Res. 13(14):673-681.
- 70. Molín J, Mendonça FS, Henderson EE, et al. (2021) Toxic wasting disorders in sheep. Animals. 11(1):229.
- 71. Muir AD. (2019) Analytical methodology for plant toxicants. In: Foodborne Disease Handbook. CRC Press. 351-412.
- Musese LJ, Andrew SM, Shirima DD, et al. (2020) Effects of Parthenium hysterophorus abundance on herbaceous species in Simanjiro, Tanzania. Int J Eng Technol Manag Res. 7(5):11-20.
- Muyt A. (2001) Bush Invaders of South-East Australia: A guide to identification and control of environmental weeds. RG & FJ Richardson. 378.
- 74. Parsons W, Cuthbertson E. (2001) Noxious Weeds of Australia. Inkata Press. 692.
- 75. Petanidou T, Godfree RC, Song DS, et al. (2012) Self-compatibility and invasiveness in native vs invasive species. Perspect Plant Ecol Evol Syst. 14(1):3-12.
- Peterson JE, Jago MV. (1984) Toxicity of Echium plantagineum: Pyrrolizidine alkaloid poisoning in rats. Aust J Agric Res. 35(2):305-315.
- Piggin CM. (1976) Factors affecting seed germination of Echium plantagineum and Trifolium subterraneum. Weed Res. 16(5):337-344.
- 78. Piggin CM. (1977) Herbaceous species of Echium naturalised in Australia. Muelleria. 3(4):215-244.
- 79. Piggin CM. (1982) Biology of Australian weeds. 8. Echium plantagineum. J Aust Inst Agric Sci. 48(1):3-16.
- Piggin C, Sheppard A. (1995) Echium plantagineum. In: Groves RH, Sheppard RCH, Richardson RG (eds) The Biology of Australian Weeds. Vol. 1. Richardson, Melbourne. pp. 87-110.
- Prasad P, Anjali P, Sreedhar R. (2021) Stearidonic acid from plants: Omega-3 source with health benefits. Crit Rev Food Sci Nutr. 61(10):1725-1737.
- Pyšek P, Jarošík V, Hulme PE, et al. (2012) Global assessment of invasive plant impacts: Interaction of traits and environments. Glob Change Biol. 18(5):1725-1737.
- Quinn JC, Kessell A, Weston LA. (2014) Plant products causing photosensitization in grazing animals. Int J Mol Sci. 15(1):1441-1465.
- 84. Rejmanek M, Richardson DM. (1996) What attributes make

some plant species more invasive? Ecology. 77(6):1655-1661.

- 85. Richardson DM, Witt ABR, Pergl J, et al. (2022) Plant invasions in Africa. In: Clements DR et al. (eds) Global Plant Invasions. Springer. 225-252.
- 86. Royal Horticultural Society (RHS). (n.d.) Echium plantagineum.
- 87. Schild AL, Motta AC, Riet-Correa F, et al. (2004) Photosensitization in cattle in Brazil. In: Poisonous Plants and Related Toxins. CABI Publ. 162-166.
- Schirmel J, Bundschuh M, Entling MH, et al. (2016) Invasive plants impact resident animals across ecosystems. Glob Change Biol. 22(2):594-603.
- 89. Seaman J, Dixon R. (1989) Toxicity of Echium plantagineum in sheep: Pen feeding experiments. Aust Vet J. 66(9):286-292.
- Seaman J, Turvey W, Ottaway S, et al. (1989) Toxicity of Echium plantagineum in sheep: Field grazing experiments. Aust Vet J. 66(9):279-285.
- Seebens H, Blackburn TM, Dyer EE, et al. (2017) No saturation in global accumulation of alien species. Nat Commun. 8(1):14435.
- 92. Shaik RS, Gurusinghe S, Weston LA, et al. (2022) Historical perspective on plant invasion in Australia. In: Clements DR et al. (eds) Global Plant Invasions. Springer, Cham. 129-149.
- 93. Shea K, Smyth M, Sheppard A, et al. (2000) Patch size and density effects of Echium plantagineum on oviposition of Mogulones larvatus. Oecologia. 124(4):615-621.
- 94. Simmonds H, Holst P, Bourke C. (2000) Palatability and toxicity of Australian weeds to goats. 1-156.
- 95. Singh V, Singh H, Sharma GP. (2011) Eco-physiological performance of invasive Ageratum species in Indo-Gangetic plains. Environ Monit Assess. 178(1):415-422.
- 96. Skoneczny D, Weston PA, Zhu X, et al. (2017) Shikonins in root periderm of invasive Echium spp. in Australia. Molecules. 22(2):330.
- Skoneczny D, Weston PA, Zhu X, et al. (2015) Pyrrolizidine alkaloid profiling in Echium spp.: A novel weapons case. Int J Mol Sci. 16(11):26721-26737.
- 98. Smith L, Culvenor C. (1981) Plant sources of hepatotoxic pyrrolizidine alkaloids. J Nat Prod. 44(2):129-152.
- Smith LM, Lonsdorf EV, Rankin EE. (2010) Invasive plants sustain ant diversity but reduce seedling recruitment. 12(1):33-43.
- 100.Sorensen TA. (1948) Method of grouping plant communities by species similarity. Biol Skar. 5:1-34.
- 101.Stout JC, Tiedeken EJ. (2017) Interactions between invasive plants and native pollinators: Impacts and responses. Funct Ecol. 31(1):38-46.
- 102.Teshome A, Abule E, Lisanework G. (2010) Rangeland use and land degradation in SE Ethiopia. Trop Grasslands. 44:202-212.
- 103.Tsegay G. (2019) Herbaceous species composition and biomass in NW Tigray under exclosure. 8(6):325-332.
- 104. Ulshöfer R, Bros H, Hauser AE, et al. (2022) Protecting axonal mitochondria from oxidative stress. Oxid Med Cell Longev. 2022:13.
- 105.Unger IM, Kremer RJ, Veum KS, Goyne KW. (2022) Short- and long-term effects of invasive plants on soil. Soil Ecol Lett. 4(3):276-288.

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- 106.Valese AC, Molognoni L, de Sá Ploêncio LA, et al. (2016) LC-ESI-MS/MS detection of pyrrolizidine alkaloids in honey. Food Control. 67:183-191.
- 107.Vilà M, Espinar JL, Hejda M, et al. (2011) Meta-analysis of invasive alien plant impacts. Ecol Lett. 14(7):702-708.
- 108. Wagstaff DJ. (2008) International Poisonous Plants Checklist: An Evidence-Based Reference. CRC Press. 464.
- 109.Wakshum S, Sebsebe D, Tamrat B. (2018) Ecological impacts of invasive alien plant species in Ethiopia. Int J Mol Biol. 3(4):171-178.
- 110.Wambua JK. (2010) Distribution and ecological impacts of invasive plants at Ol-Donyo Sabuk NP, Kenya. MSc Thesis Univ Nairobi Kenya. 118.
- 111.Wang W, Jin J, Xu H, Shi Y, Boersch M, Yin Y. (2022) Comparative analysis of medicinal substances in Echium vulgare and Echium plantagineum: A review. J Ethnopharmacol.

285:114894.

- 112.Weber E. (2017) Invasive Plant Species of the World, 2nd edn: A Reference Guide to Environmental Weeds. CABI.
- 113.Weston PA, Weston LA, Hildebrand S. (2013) Metabolic profiling of Echium plantagineum from southeastern Australia. Phytochem Rev. 12(4):831-837.
- 114.Witt A, Luke Q (eds). (2017) Guide to the Naturalized and Invasive Plants of Eastern Africa, 1st edn. CABI. 609 p.
- 115.Zhu X, Meyer L, Gopurenko D, et al. (2014) DNA barcoding for identifying two Echium invaders in Australia. 5.
- 116.Zhu X, Skoneczny D, Weidenhamer JD, et al. (2016) Bioactive naphthoquinones in Echium plantagineum rhizosphere. J Exp Bot. 67(12):3777-3788.
- 117.Zhu X, Weston PA, et al. (2017) Ecology and genetics of Echium species invasion success in Australia. Sci Rep. 7(1):42792.